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Clonidine Plus GHRH Administration for Diagnosing Growth Hormone Deficiency in Children

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Abstract

Objective: To analyze the effectiveness and reproductivity of a challenge with clonidine (CLO) plus GHRH administration to establish a state of GH-deficiency and the causes of it.

Methods: 24 children (17M, 7F) with stature <1 or 2 SD from P3, previously studied with different classical tests, were given CLO (150 µg/m2 orally, at time 0') plus GHRH (60' later, 1 μg/kg, iv). This test was repeated but giving placebo at time 0'. For comparison purposes, 20 agematched normal children (14M, 6F) were studied. Growth velocity (GV) and bone age (BA) before performing CLO +GHRH were considered.

Results: While classical GH tests show a great variability of negative or positive GH responses in the same short patient, all but two patients significantly responded to CLO+GHRH (GH peak: 42.2 ± 19 vs. 41.6 ± 3.5 in controls). Controls showed a significant positive correlation between the amplitude of GH response and chronological and bone age (R=0.832; F=37.6; P=0.001, multiple regression analysis): y=a+b × BA+c × CA, where y=GH peak, BA=bone age, CA=chronological age, and a, b and c are constants. This equation allows to establish the value of GH peak theoretically obtained after CLO+GHRH challenge in a normal child. Applying this equation to the responses to CLO+GHRH in short children a deviation, ranging between -435% and +41%, was obtained between theoretical and real GH peaks (%DT-R). Responses between -20% to +20% were considered normal. Plotting these responses (% DT-R) against the SD of the mean of the BA to GV ratio in normal children, five populations were defined: 1) Low response and low GV for BA; 2) Low response and normal GV for BA; 3) Normal response and low GV for BA; 4) Normal response and normal GV for BA; and 5) High response and low GV for

Conclusion: The test CLO+GHRH is safe, effective and unique to detect GH-deficiency and its causes in short children and adults.

Keywords: Growth hormone; Clonidine; Growth hormone releasing hormone; Growth hormone-deficiency; IGF-I; Bone age; Hypothalamic-somatotroph rhythm.

Introduction

Since the discovery in 1982 of growth hormone releasing hormone (GHRH) by Vale et al. [1] and Guillemin et al. [2], and the availability of this peptide, a number of studies demostrated that it was a specific inducer of GH secretion in man. On this basis it was tried to establish its utility as a diagnostic useful tool for clearly defining when a child was GH-deficient (GHD), and even more it was thought that this peptide would allow to define whether the deficiency of GH secretion occurred as a consequence of an hypothalamic dysfunction, or affectation, or it has its origin at the pituitary level [3-6]. Moreover, it was thought that in the cases of GHD due to a hypothalamic dysfunction the administration of GHRH might replace the treatment with GH [7-9].

However, physiologically, pituitary GH secretion is mainly subjected to a negative control exerted by somatostatin (SS), released from the hypothalamus into the portal blood; SS also negatively regulates hypothalamic GHRH secretion. Therefore, only when there is a physiological interruption of SS secretion, GHRH may be released and act on the pituitary synthesis and release of GH [10,11].

We demonstrated the existence of an endogenous hypothalamic-somatotroph rhythm (HSR) in man [12], responsible for the high variability in the GH responses to a GHRH test, even in the same individual tested at different daytimes.

According to our postulate, the administration of GHRH during a refractory phase of HSR, would not induce a significant GH response (GH peak<10 ng/ml), since during this phase hypothalamic SS would be physiologically released, therefore blocking the effect of GHRH on somatotrophs. This is the main reason by which most of GHRH tests carried out for analyzing whether GH-deficiency exists in short stature children may give false negative results. In that study we demonstrated the need of taking into account plasma GH values (-30, -15, 0 minutes)

prior to GHRH challenge for knowing the HSR phase at which the test was performed and hence avoiding erroneous interpretation of the results obtained [12]. Most likely the lack of consideration of this concept led to GHRH being discontinued as a single test for analyzing GH-deficiency.

Clinically, the finding of a test that would allow to clearly defining when GH-deficiency exists and the origin of it would be very useful. Moreover, this putative test would also allow to decide when a treatment with GH is absolutely necessary and when this treatment should start. Many tests for analyzing GH secretion have been developed (propranolol plus exercise, levodopa, arginine, glucagon, insulin hypoglycemia, etc) [13-16], but perhaps because of the complexity of the mechanisms involved in GH neuroregulation, still they are controversed; mainly because of the lack of reproductibility, but also because most of them can not differentiate whether there is a pituitary affectation leading to insufficient or deficient GH secretion, or the affectation is due to a hypothalamic dysfunction. In addition, these classical tests are not able to differentiate between a neurosecretory dysfunction (transient or chronic), or when there is a Constitutional Growth Delay or Short Family stature, especially when Short Family stature is associated to a genetic reason. In addition, some of them may imply risks for the health of the child being analyzed, as it happens with the hypoglycemia induced by insulin (specially contraindicated in children with brain injuries, epilepsy or ischemic cardiopathy) since glycemia has to decrease about to 40 mg/dl for achieving a clear result in terms of GH response. In fact, some deaths occurred during this test, and recently we knew that the test had to be early interrupted since glycemia was as low as 20 mg/dl.

As stated above, it was thought that GHRH would be very useful to establish and characterize GH-deficient states. However, it was not happened; in general, and regardless of the etiology of GH-deficiency, most of the patients tested with GHRH showed a GH response to this stimulation [17]. Perhaps this was due to the fact that a hypothalamic dysfunction (absence or inadequated GHRH secretion into the portal blood) is the responsible reason for the lack of a physiological spontaneous GH release. On the other hand, subjects which did not respond to GHRH test could have been challenged during a refractory phase of the hypothalamic somatotroh rhythm [12], not taken into account when analyzing the GH responses to GHRH. This can occur either physiologically (normal trough period), or pathologically if there is a chronically increased somatostatinergic tone. Therefore, the lack of utility of GHRH as a tool for analyzing whether GH-deficiency exists led to discard its use for this purpose.

Hypothalamic somatostatin release is negatively controlled by adrenergic and cholinergic pathways [10,11]. Therefore, the administration of alpha2-adrenergic agonists such as clonidine (CLO) will reduce, or impede, somatostatin secretion, allowing pituitary GH release to be enhanced [18-20]. The same occurs with the cholinergic agonist pyridostigmine [21], or galanin, which acting through the cholinergic pathways increases the GH response to GHRH in normal man [22,23]. On this basis it was postulated that the test Pyridostigmine plus GHRH would allow establishing and differentiating a deficient GH secretion of

pituitary origin from one due to a hypothalamic dysfunction [24], without establishing whether there could exist a Congenital Delay of Growth or Short Family stature. Moreover, the fact that alpha2-adrenergic receptor blockade is able to blunt the increased response to pyridostigmine plus GHRH [25] indicates that this Pyridostigmine plus GHRH test may lead to false negative responses (indicating a false state of GH-deficiency). On the contrary, pre-treatment with CLO increases the GH response to GHRH in short and normal children and also in adults [18,26], but it was not known whether such a test could provide a differential diagnosis, and even treatment, of the diverse situations of GH-deficiency, or insufficiency in children.

In 1993, we tried to give an answer to the issues raised above performing a study, in which we compared the GH responses to the combined administration of CLO plus GHRH in children with stature below P3 (previously studied for GHD with other tests), with those obtained after this CLO plus GHRH stimulation in a population of normal children, matched in age and sex. Our data indicated that the CLO+GHRH test is the most powerful test for establishing when a GHD exists and the causes leading to it.

Patients and Methods

Short stature children

24 children (17 boys and 7 girls, aged 7 to 13.9 years) with stature below one or two standard deviations from the third percentile (P3), participated in the study after obtaining signed informed consent from their parents or their legal representatives. The procedures used were in accordance with the guidelines of the Helsinki Declaration of human experimentation, and the study was approved by the Ethics Committee of the School of Medicine (University of Santiago de Compostela).

The weight of 7 of these children was equal or lower or below P3, and in 3 of them there was a clear malnourishment (Body Mass Index<14 kg/m²). All of them were in pre pubertal status (Tanner I-III). Two children had received prophylactic cranial irradiation due to brain tumors, 5 and 3 years before the study. One of the children had suffered a traumatic brain injury 3 years before. 12 children had been considered to have Short Family stature. In 4 of the cases existed low weight at birth, while the remaining children had been considered to present Constitutional Growth delay. All children had been previously studied for their GH deficiency or insufficiency: growth velocity, bone age, classical provocative tests for GH secretion (i.e. clonidine, insulin hypoglycaemia and propranolol plus exercise), plasma Insulin Growth Factor-I (IGF-I) analysis, and analysis of the spontaneous pattern of nocturnal GH secretion (area under the GH curve [AUC] in blood samples drawn at 20-minutes intervals during the night.

Studies for analyzing the GH response to CLO+GHRH commenced at 09.00 h after an overnight fast and bed rest and thirty minutes after the insertion of an indwelling catheter for blood withdrawal in a forearm vein. The study consisted of administering CLO (Catapresan, Boehringer Ingelheim, Spain) orally, 150 $\mu g/m^2$ at time 0 minutes, followed by the

administration of GHRH (GRF 1-29, Serono, Spain) as an intravenous bolus (1 $\mu g/kg$) at time 60 minutes. Three days later, an additional control for short children was performed, administering placebo at time 0, and then GHRH 60 minutes later.

Control children

In 20 age-matched normal stature children (14 boys and 6 girls) the GH responses to CLO plus GHRH were analyzed, as in short stature children, after evaluating the growth velocity during six months and the bone age (X-ray of the left wrist).

In both groups of children, blood samples for GH analysis were drawn at 0 and 60 minutes and at 15 minute intervals for one hour following GHRH administration. Plasma GH was measured by radioimmunoassay using a commercial kit (BioMerieux, France). The mean intra-assay coefficient of variation was 5.7, 3.2 and 4.3% at mean GH concentrations of 1.5, 10 and 22 ng/ml respectively. To avoid interassay variations, all samples were run in the same assay, after being stored at -4°C until assayed.

Statistics

To establish the normal range of GH peaks in response to CLO +GRF in normal children a multivariate regression analysis was performed. The non-parametric Wilcoxon's test was used to

compare the results obtained in the CLO+GRF test in both group of children. A value of P<0.05 was considered to be statistically significant. Data are presented as Mean \pm SEM, unless other parameters are indicated.

Results

Short stature children

56% of these children showed a GH response to CLO<10 ng/ml, the same happened in 36% of children after being stimulated with propranolol plus exercise, and in 6 of the 7 patients studied with the insulin-induced hypoglycemia (Table 1). However, in all but 6 children, indices of total and pulsatile nocturnal GH release appeared to be normal; plasma IGF-I values were below normal limits for age in 18 of them. Mean \pm SEM plasma IGF-I was 51 \pm 7.1 ng/ml (normal range for the age: 47-315 ng/ml). Mean \pm SEM growth velocity was 3.7 \pm 0.9 cm/yearr. Thyroid hormones and cortisol were in normal values (data not shown).

GHRH administration 60 minutes after placebo, induced a GH peak of 19.5 ± 15.7 ng/ml. This value was significantly (P<0.01) increased when GHRH was administered 60 minutes after CLO administration: 42.2 ± 19 , although two children did not show any significant change in the GH peak after CLO+CHRH challenge (GH peak < 10 ng/ml).

Table 1: Number of positive (+, > 10 ng/ml) or negative (-, < 10 ng/ml) GH peaks in response to different provocative tests performed in Short Children. Note that this group of children has been subdivided into five different groups according to the individual deviations between the theoretically calculated GH peak response to CLO+GHRH and the real value obtained. (See Table 2 for a better understanding of this concept). Note the great variability in the responses to different stimuli in each subgroup.

Subgroup	n	P+E	CLO	IH	GHRH	CLO+GHRH
1	6	+3, -3	+0, -6	+0, -6	+2, -4	+4, -2
2	5	+3, -2	+4, -1	NP	+3, -2	5
3	5	+3, -1	+2, -2	NP	+3, -1	5
4	5	+4, -1	+3, -2	NP	+3, -2	5
5	3	+1, -1	+1, -2	1	3	3

n=number of children in each subgroup. P+E=challenge with propranolol plus exercise. CLO=stimulation with oral clonidine. IH=Insulin-induced hypoglycemia. NP= not performed.

Control children

Pre-treatment with CLO led to GH response to GHRH of 41.6 ± 3.5 ng/ml, a peak value not significantly different to that obtained in short stature children after CLO+GHRH test. There was a significant positive correlation between the amplitude of GH response and both chronological and bone age (R=0.832; F=37.6; P=0.001), as established by a multiple regression. This analysis gave origin to an equation: $y=a+b \times BA+c \times CA$, where y=GH peak, BA=bone age, CA=chronological age, and a, b and c are constants obtained. Values of a, b and c can vary according to the number of subjects in the normal population analyzed. This positive correlation was also observed between GH peaks and the bone age/growth velocity ratio in this group of normal children. Therefore, this equation would allow establishing the

value of GH peak that theoretically should be obtained after CLO +GHRH challenge in a normal stature child.

To confirm the validity of CLO plus GHRH test to establish GH-deficient or insufficient states and to differentiate among their causes, we calculated the theoretical GH responses to this test in short children according to the equation above described. There was a deviation between theoretical values of GH peaks which should be obtained with this equation and the real GH peak values obtained after CLO plus GHRH. The percentage of the deviation between theoretical values (T) and real values (R) ranged between -435% and +41% (Table 2). Responses between the -20% to +20% range were considered to be normal, as they fitted between one standard deviation from the mean in the group of normal children. Plotting these responses (% DT-R)

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against the standard deviation of the mean of the bone age to growth velocity ratio in normal children, five populations were showed to exist: 1) Low response and low growth velocity for bone age; 2) Low response and normal growth velocity for bone age; 3) Normal response and low growth velocity for bone age; 4) Normal response with normal growth velocity for bone age; 5) High response with low growth velocity for bone age. These data are shown in Figure 1.

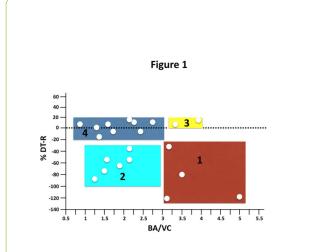


Figure 1: Differential diagnosis of GH-deficiency according to the percentage of deviation between the theoretical GH peak induced by CLO+GHRH challenge and the real peak obtained after this stimulation (y axis, % DT-R) in short stature children, plotted against the deviation of the bone age to growth velocity ratio in normal children (x axis, BA/VC). % DT-R values comprised between -20 and 20 are considered as normal responses (see the text), while values over 20 are considered as high responses and values below -20 are considered as low responses. 1 (red rectangle): indicates pituitary GH deficiency or insufficiency. 2 (light blue rectangle) indicates Neurosecretory Dysfunction. 3 (yellow rectangle): indicates Short Family stature. 4 (dark blue rectangle): indicates Constitutional Growth Delay. In this Figure the subgroup 5 is not represented because of the low body mass index and deficient nutritional conditions existing in the three children of this subgroup.

Discussion

According to our results it seems to be clear that the varied situations of GH-deficient or insufficiency may be differentiated by the combined administration of an alpha-2-adrenergic agonist, such as clonidine, and GHRH. This test both analyzes the functionality of the hypothalamic zones involved in GH control, and directly evaluates the secretory integrity of somatotropes. Therefore, no false negative responses are expected to occur. Moreover, it minimizes, at high GH levels, the variability in GH responses to GHRH, thus allowing its use in the treatment of short children having a hypothalamic dysfunction responsible for their diminished linear growth.

Therefore, it seems that the CLO plus GHRH test has a strong and an unique value to allow better differentiation among the diverse GH-deficient states; therefore it might optimize the decision about which children must be treated and when the treatment must start.

Diagnosis of GHD is based on auxologic, clinical and hormonal criteria [27-29]. The latter is mainly related to the GH response to provocative tests. However, until now, all of the known provocative tests for GH secretion usually lead to a number of false negative results. This is the main reason by which Regulatory Authorities and GH-Committees established that for improving the validity of the biochemical diagnosis of GHD two different provocative stimuli have to be performed. Nevertheless, results here presented (Table 1) are a clear demonstration that a same child may respond negatively or positively to two different challenges. Interestingly, in that Table 1 it could be seen that none of the children of the subgroup 1 responded to the Insulin-induced hypoglycaemia, a test considered to be the "gold standard test", while four of them showed a positive response to CLO + GHRH, and even in two of these patients a response to GHRH given alone could be observed. This is quite surprising since it is well known that hypoglycaemia induces an adrenergic response; in fact catecholamines form part of the group of counter regulatory hormones that oppose the action of insulin, and they also act by blocking hypothalamic somatostatin release [18], then allowing the endogenous GHRH to be released [10,11]. Therefore a positive response should be observed in these cases, as it happened in four of them when they were tested with CLO + GHRH or in three of these children after being stimulated with propranolol plus exercise (Table 1). The only feasible explanation for these divergences is the phase of HSR at which they had been tested [12], not considered when the stimulation was performed. Another possibility, such as the lack of hypothalamic GHRH production, has to be discarded since in this subgroup 1 there was a positive response to propranolol plus exercise in three cases, and in two patients challenged with GHRH alone (Table 1). Since GHRH induces the pituitary synthesis of GH [10,11], no response to exogenous GHRH administration should be observed in these two patients given that no pituitary stores of GH could exist. These divergences among the different tests carried out were also observed in the other subgroups of children, with the unique exception of CLO+GHRH test for which a positive GH response was observed in all cases (Table 1).

Apart of this, the cut-off value to establish whether a patient suffers GH-deficiency is quite arbitrary for most of the tests usually utilized and it has been changing during years. In addition, classical provocative tests are unable to characterize the various GH-deficient states. Therefore, the utility of these tests continues to be a matter of controversy [30]. In fact, more than 30 different stimuli for analyzing GH secretion have been already described [31]. As a clear consequence of these controversies a survey carried out in USA among paediatric

endocrinologists concluded that, in general, main indications to treat a paediatric patient with GH are: poor growth velocity, poor target height and low plasma IGF-I levels [32].

Table 2: Percentage of deviation (%DT-R) between the theoretically calculated GH peak response to CLO+GHRH challenge (according to the equation $y = a + b \times BA + c \times CA$) and the GH peak obtained after this stimulation in each subgroup of Short Children. Note that the highest deviation was observed in the subgroup 1. Results are shown as the Mean \pm SEM, but the range of deviations oscillated between -435% and +41%.

Subgroup	n	%DT-R	GV	мсен	CA	ВА
1	6	211 ± 163	2.8 ± 0.06	4.1 ± 2.1	10.9 ± 2.1	8.3 ± 2
2	5	-69 ± 36	4.5 ± 0.4	5.2 ± 0.9	11 ± 1.3	8.1 ± 1.7
3	5	9 ± 6.2	3.6 ± 0.4	4.9 ± 2	8.4 ± 1.2	5.9 ± 1.7
4	5	4 ± 9	4.4 ± 1	5.9 ± 1	11 ± 2.2	8.4 ± 3
5	3	36 ± 5	3.3 ± 0.3	7.8 ± 1.8	8.3 ± 1.2	4.6 ± 1.5

GV=Growth velocity (cm/year). MCGH=Mean ± SEM plasma GH concentrations (ng/ml) during a nocturnal period of sampling. CA=Chronological age (years). BA=Bone age (years).

That is, tests of GH secretion in short stature children are not considered priorities for commencing a treatment with the hormone. At this point and although it is not the objective of this study, low plasma IGF-I values do not necessarily reflect GH-deficient secretion. As we recently published, the hepatic expression of IGF-I is strongly conditionated by the intracellular metabolism of glucose [33], therefore poor nutritional conditions may lead to low plasma levels of this factor despite of normal or increased GH secretion, as it happens in Anorexia Nervosa patients [34]. It can occur too in children with strong spasticity due to brain injuries. The high avidity of spastic muscles for glucose may lead to low IGF-I production by the liver.

In summary, in this study carried out in 1993, we demonstrated that the test CLO+GHRH is useful for analyzing GH a posible GH-deficiency in short stature children and establishing the most probable causes of it. According to our results, the different subgroups of short children analyzed could be classified into other 5 groups (Figure 1): children from Group 1 are GH-deficient, from pituitary origin in two of them, and hypothalamic (neurosecretory dysfunction) in the other two. While the first two would to be treated with GH, the other two would benefit from a treatment with CLO+GHRH. Children from Group 2 suffer a Neurosecretory Dysfunction, therefore they also would need GH treatment although they could also be treated with CLO+GHRH. In Group 3 we placed 2 children with Short Family stature; most likely GH treatment or CLO+GHRH would increase their growth velocity and height. The 9 children in Group 4 appear to have Constitutional Growth Delay; therefore no GH therapy would be needed in this group, although gonadal function should be studied in them. Lastly, children from Group 5 were malnourished; hence pre-treatment with clonidine did not significantly enhance the GH response to GHRH.

In summary, the combined test with clonidine and GHRH allows the differentiation among diverse GH-deficient states. Pre-treatment with CLO places the subjects in a similar basal

state thus eliminating interferences due to the hypothalamic-somatotroph rhythm [12]. Therefore, GH responses would be a real expression of the pituitary reserve of GH, indicating the functionality of the hypothalamus-somatotrope axis. Hence, this test does not give false negative responses and facilitates the differential diagnosis of GH-deficiency without the need for other provocative tests or the analysis of spontaneous GH secretion. In addition, some children growing below P3 and showing a positive response to CLO+GHRH, independently of the degree of deviation from theoretically calculated values might benefit from a treatment with both compounds.

For carrying out the CLO+GHRH test and analyzing the GH responses in short children, a normal population of different prepubertal ages should be studied before to establish the equation $y = a + b \times BA \times c \times CA$, which would serve as reference for the normality or not of the GH peak. Even more, the normal responses to this test should be separately analyzed in boys and girls. Unfortunately, GHRH is no longer available in many countries (including Spain). Moreover, since we developed this test 24 years ago, a number of factors that may interfere with the GH responses have been discovered. Among them, the intricate relationships between auto/paracrine effects of GH, SS and GHRH on its own secretion, the pituitary production of IGF-I, the role of the gastric hormone Ghrelin, or the release of leptin by adipocytes, nutritional conditions, stress, etc. [11]. In addition, many recent data indicate that the aging-suppressor gene Klotho is a direct regulator of GH secretion [35]. However, we still think that the CLO+GHRH test is a clear reproducible and safe test for analyzing possible states of GH-deficiency and its causes, even in adulthood [36].

Conflict of Interests

No conflicts of interest exist.

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